

The potency of both compounds in the inhibition of enzyme in-vitro is of the same magnitude or greater compared with those of the  $H^+$ ,  $K^+$ -ATPase inhibitors omeprazole (Im et al 1985) and SCH 28080 (Scott & Sundell 1985) which exhibited  $IC_{50}$  values of 6.0 and 2.5  $\mu M$  in our assay system, respectively. The antisecretory and anti-ulcer activity in-vivo, however, was less effective. II, in particular, was inactive in doses up to 200 mg  $kg^{-1}$ , p.o. for inhibiting gastric lesions. One possible explanation for the discrepancy observed in-vitro and in-vivo is the poor hydrophobicity of the compounds. Since these compounds act at the cytosolic site after absorption, a poor hydrophobicity may prevent them from reaching the  $H^+$ ,  $K^+$ -ATPase site. Also, the possibility that these compounds may be readily degraded to inactive metabolites cannot be ruled out.

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*J. Pharm. Pharmacol.* 42: 726-728  
Communicated April 13, 1990

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## Comparison of intestinal and peritoneal dialysis of theophylline and phenobarbitone in rats

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**Abstract**—Intestinal dialysis of drugs by oral administration of activated charcoal has been compared with peritoneal dialysis in rats. The average amounts of theophylline transported over 120 min into the intestinal lumen and the peritoneal cavity were 15.7 and 16.5% of the intravenous dose (10 mg  $kg^{-1}$ ), respectively, showing no significant difference, whereas the amount of the same intravenous dose of phenobarbitone transported from the blood into the intestinal lumen (7.8%) was significantly smaller than that entering the peritoneal cavity (12.5%). The net water flux showed that secretion predominated in the peritoneal transport whilst absorption predominated in the intestinal transport for both drugs. However, the net water flux in the intestinal lumen after intravenous theophylline (as aminophylline) was significantly smaller than that following phenobarbitone. The differences in transport across the two membranes could be due to differences in the intrinsic properties of the membranes, such as the surface area, the thickness of the membrane and the distribution of blood vessels. Differences could also be due to differences in the pharmacological effects of the drugs.

In acute drug overdoses, haemoperfusion, haemodialysis, peritoneal dialysis, and combined haemodialysis-haemoperfusion have been used as methods of haemopurification (Weinberger & Hendeles 1980; Gibson 1981). Oral administration of activated charcoal has also been employed to inhibit absorption of excess drugs from the gastrointestinal (GI) lumen into the blood. The GI mucous membranes have a large surface area when considered as a whole. In particular, the total absorptive or exsorptive area of the small intestine has been calculated to be about 200  $m^2$  in an adult human, and is far larger than that of the peritoneal membrane (about 2  $m^2$ , Csaky 1984).

We have previously reported that intravenously administered drugs are generally transported into the small intestinal lumen and the bile in rats, and that drugs can be removed by adsorption onto orally administered activated charcoal (Arimori & Nakano 1986a, 1987, 1988a, b). However, GI dialysis by oral administration of activated charcoal has not been clearly established as an effective means of removing drugs which have been parenterally administered or have already been absorbed into the systemic circulation from the GI tract. The present study compared GI

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dialysis with peritoneal dialysis, which has been established as a valuable means of haemopurification in drug overdoses, for theophylline and phenobarbitone.

### Materials and methods

**Materials.** Aminophylline (Neophyllin) and theophylline were purchased from Eisai Co. (Tokyo) and Wako Pure Chemical Industries (Osaka), respectively. Phenobarbitone sodium was obtained from Daiichi Seiyaku Co. (Tokyo). Scintillation fluid (Instagel) was from United Technologies (Packard, Illinois, USA). All other chemicals used were of analytical grade.

**Intestinal dialysis.** Male Wistar rats, 220–360 g, were fasted overnight with free access to water. Intestinal exsorption experiments were performed using an in-situ single-pass perfusion technique (Arimori & Nakano 1985). Lactated Ringer solution (37°C) was perfused at 1.3 mL min<sup>-1</sup> from the duodenum through the small intestine to the ileocaecal junction using a perfusion pump. Theophylline (as aminophylline) or phenobarbitone was injected over about 1 min at a dose of 10 mg kg<sup>-1</sup> into the right femoral vein. Blood samples and dialysate were collected periodically.

**Peritoneal dialysis.** A small incision was made in the abdomen and 20 mL of lactated Ringer solution (37°C) was injected into the peritoneal cavity. The dialysate was exchanged for the corresponding volume of new dialysate every 15 min. In the control group of rats, the same operation was performed without injection of the dialysate. Administration of drugs and collection of samples were performed as above.

**Water flux.** The rate of water movement from the blood into the GI lumen or peritoneal cavity and from the lumen or peritoneal cavity back to the blood was calculated from changes of concentrations in <sup>3</sup>H<sub>2</sub>O (0.74 MBq) and phenol red (10 µg mL<sup>-1</sup>), used as a non-absorbable marker.

**Analytical methods.** Theophylline and phenobarbitone in the serum and dialysate were determined by high-pressure liquid chromatography (Arimori & Nakano 1985, 1986b). Phenol red was determined spectrophotometrically by absorbance at 550 nm after addition of 4 mL 1M NaOH to 0.5 mL of dialysate. For the assessment of <sup>3</sup>H<sub>2</sub>O concentrations, 5 mL of Instagel scintillation fluid was added to 200 µL of dialysate and radioactivity determined in a liquid scintillation counter.

**Pharmacokinetic analysis.** The intestinal and peritoneal clearance values of theophylline and phenobarbitone were calculated by dividing the overall amount of the drugs transferred into both dialysates in 120 min by the appropriate value for an area under the serum concentration-time curve of the drugs, obtained over the same period of time. The unpaired *t*-test was used to evaluate significant differences.

### Results and discussion

Fig. 1 shows the transfer rate of theophylline and phenobarbitone from the blood into the GI lumen and the peritoneal cavity following intravenous administration (10 mg kg<sup>-1</sup>). The transfer rate of theophylline into the intestinal lumen was similar to that into the peritoneal cavity, with the average amounts of the drug transported into the intestinal and peritoneal dialysates in 120 min 15.7 and 16.5%, respectively. The transfer rate of phenobarbitone into the lumen was significantly lower than that into the peritoneal cavity (*P* < 0.01); the average amounts of the drug transported into the intestinal and peritoneal dialysates in 120 min were 7.8 and 12.5%, respectively.

The intestinal and peritoneal clearances of theophylline were 33.8 and 28.0 mL h<sup>-1</sup>, respectively, and were not significantly different while the intestinal and peritoneal clearances of phenobarbitone were 12.7 and 22.6 mL h<sup>-1</sup>, respectively.

Table 1 shows water movement into the GI lumen and peritoneal cavity after intravenous administration of aminophylline or phenobarbitone at a dose of 10 mg kg<sup>-1</sup>. Water movement from the blood into the lumen was less than that from the blood into the peritoneal cavity in both cases. The net water flux showed that secretion predominated in the peritoneal cavity, while absorption predominated in the lumen for both drugs. However, the net water flux into the lumen after intravenous administration of theophylline was significantly smaller than that following phenobarbitone.

Differences in transport across each of the membranes could be due to differences in their intrinsic properties, such as the surface area, the thickness of the membrane, and distribution of blood vessels. They could also be due to the different pharmacological effects of the drugs. The small intestine has a much larger surface area than the peritoneal membrane and therefore the intestine would be expected to be a more permeable or exsorbable organ compared with the peritoneum. Published studies have demonstrated that the net water flux causes a drag effect on permeability of some drugs (Ochsenfahrt & Winne 1974a, b; Kitazawa et al 1975, 1977; Karino et al 1982a, b).

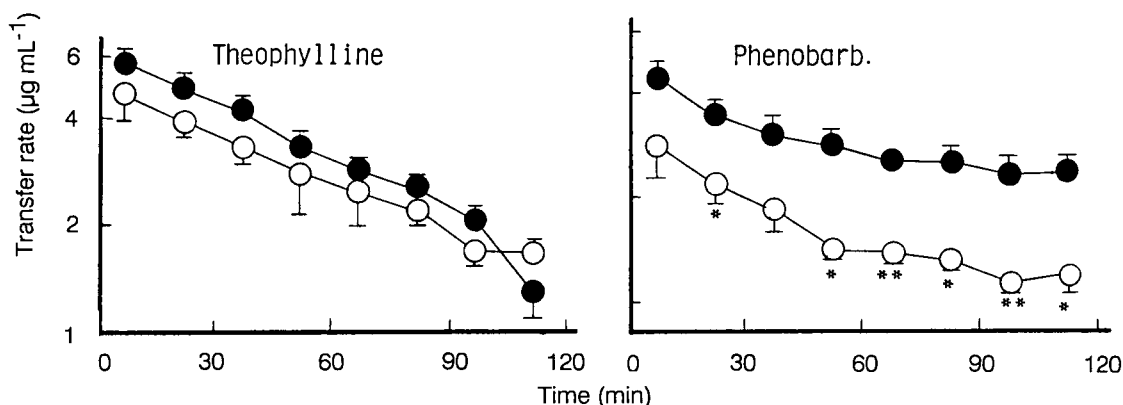


FIG. 1. The transfer rate of theophylline and phenobarbitone from the blood into the intestinal lumen (○) and the peritoneal cavity (●) after intravenous administration of theophylline (as aminophylline) and phenobarbitone (10 mg kg<sup>-1</sup>) to rats. Each point represents the mean ± s.e.m. of 4 rats. \**P* < 0.05, \*\**P* < 0.01.

Table 1. Water flux into the intestinal lumen or the peritoneal cavity after intravenous administration of theophylline (as aminophylline) and phenobarbitone (10 mg kg<sup>-1</sup>).

	Water flux from blood to intestinal lumen or peritoneal cavity (mL h <sup>-1</sup> )	Water flux from intestinal lumen or peritoneal cavity to blood (mL h <sup>-1</sup> )	Net water flux <sup>a</sup> (mL h <sup>-1</sup> )
Intestinal lumen			
Theophylline	34.4 ± 0.32	34.5 ± 1.00	-0.10 ± 1.29
Phenobarbitone	35.9 ± 3.80	39.6 ± 4.64	-3.67 ± 0.90 <sup>d</sup>
Peritoneal cavity			
Theophylline	49.6 ± 0.32 <sup>b</sup>	44.1 ± 1.27 <sup>b</sup>	+5.47 ± 1.55 <sup>c</sup>
Phenobarbitone	53.8 ± 1.31 <sup>b</sup>	48.0 ± 1.65	+5.81 ± 0.83 <sup>b</sup>

Each value represents the mean ± s.e.m. of 5 rats. <sup>a</sup>Positive (+) signs indicate secretion and negative (-) signs indicate absorption. <sup>b</sup>*P* < 0.01, <sup>c</sup>*P* < 0.05 vs respective drug in intestinal lumen. <sup>d</sup>*P* < 0.05 vs theophylline in intestinal lumen.

For example, Kitazawa et al (1975) reported that in rats absorption of sulphanilamide, sulphisoxazole, and metoclopramide increased with increasing transmucosal fluid movement from the lumen to the blood and decreased when the movement of water was directed from the blood to the lumen.

During perfusion with lactated Ringer solution in the present study, the net water fluxes were from the GI lumen to the blood and from the blood to the peritoneal cavity (Table 1). This indicates that the effect of solvent drag upon permeability of the intestinal membrane is less than that on permeability of the peritoneal membrane and may be one of the reasons why the amount of phenobarbitone transported from the blood into the GI lumen was less than that entering the peritoneal cavity.

The greater exsorption of theophylline into the GI lumen may be due to the pharmacological effects of this drug. It has been shown that theophylline causes net water secretion by increasing mucosal cyclic AMP levels via phosphodiesterase inhibition (Waldman et al 1977; Beubler 1980). As shown in Table 1, with theophylline, the net intestinal water flux (or the net water absorption from the lumen to the blood) was smaller than with phenobarbitone, although intravenous administration of aminophylline (10 mg kg<sup>-1</sup>) did not cause net water absorption to convert into net secretion. Therefore, theophylline produces an environment in which it is easier for the drug to enter the GI lumen. Accordingly, the GI lumen may have a more noticeable exsorptive function in a theophylline overdose compared with other drug overdoses.

More data on the transport of other drugs are needed to better compare the efficacy of removal by intestinal and peritoneal dialysis.

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